RIFAMPICIN, ISONIAZID, PYRAZINAMIDE AND ETHAMBUTOL HYDROCHLORIDE TABLETS
Final text for addition to The International Pharmacopoeia

This monograph was adopted at the Fortieth WHO Expert Committee on Specifications for Pharmaceutical Preparations in October 2005 for addition to the 4th edition of the International Pharmacopoeia.

Category. Antituberculosis drugs.

Storage. Rifampicin, Isoniazid, Pyrazinamide and Ethambutol hydrochloride tablets should be kept in a tightly closed container, protected from light.

Additional information. Strength in the current WHO Model list of essential medicines: 150 mg Rifampicin, 75 mg Isoniazid, 400 mg Pyrazinamide and 275 mg Ethambutol hydrochloride. The tablets are coated.

Requirements

Comply with the monograph for “Tablets”.

Definition Rifampicin, Isoniazid, Pyrazinamide and Ethambutol hydrochloride tablets contain Rifampicin, Isoniazid, Pyrazinamide and Ethambutol Hydrochloride. They contain not less than 90.0% and not more than 110.0% of the amounts of rifampicin (C_{43}H_{58}N_{4}O_{12}), isoniazid (C_{6}H_{7}N_{3}O), pyrazinamide (C_{5}H_{5}N_{3}O) and ethambutol hydrochloride (C_{10}H_{24}N_{2}O_{2}, 2HCl) stated on the label.

Manufacture The manufacturing process and the product packaging are designed and controlled so as to minimize the moisture content of the tablets. They ensure that, if tested, the tablets would comply with a loss on drying limit of not more than 30 mg/g when determined by drying freshly powdered tablets to constant mass under vacuum at 60°.

Identity tests

- Either tests A and B or test C may be applied.

A. See Assay method A described below. The retention times of the three principal peaks in the chromatogram obtained with solution (1) correspond to those of the principal peaks in the chromatogram obtained with solution (2).
B. See the test described below under “Assay method B”. The retention time of the principal peak in the chromatogram obtained with solution (1) corresponds to that of the principal peak in the chromatogram obtained with solution (2).

C. Carry out test C.1. or, where UV detection is not available, test C.2.

C.1. Carry out the test as described under 1.14.1 Thin-layer chromatography, using silica gel R6 as the coating substance and a mixture of 100 volumes of methanol R and 1.5 volumes of strong ammonia solution R as the mobile phase. Apply separately to the plate 5 µl of each of the following two solutions in methanol R. For solution (A) shake a quantity of the powdered tablets equivalent to about 5 mg Isoniazid for 15 minutes with 5 ml of methanol R, filter, and use the filtrate. For solution (B) use 1 mg isoniazid RS and proportional quantities (according to the ratio in the tablet) of rifampicin RS, pyrazinamide RS and ethambutol hydrochloride RS per ml of methanol R. After removing the plate from the chromatographic chamber, allow it to dry in a current of air, place in a chamber with iodine vapours, and allow to stand for 20 minutes. Examine the chromatogram immediately in ultraviolet light (254 nm).

The principal spots obtained with solution A correspond in position, appearance and intensity to those obtained with solution B.

C.2. Carry out the test as described under 1.14.1 Thin-layer chromatography, using silica gel R5 as the coating substance and a mixture of 100 volumes of methanol R and 1.5 volumes of strong ammonia solution R as the mobile phase. Apply separately to the plate 5 µl of each of the following two solutions in methanol R. For solution (A) shake a quantity of the powdered tablets equivalent to about 5 mg Isoniazid for 15 minutes with 5 ml of methanol R, filter, and use the filtrate. For solution (B) use 1 mg isoniazid RS and proportional quantities (according to the ratio in the tablet) of rifampicin RS, pyrazinamide RS and ethambutol hydrochloride RS per ml of methanol R. After removing the plate from the chromatographic chamber, allow it to dry in a current of air, place in a chamber with iodine vapours, and allow to stand for 20 minutes. Examine the chromatogram immediately in daylight.

The principal spots obtained with solution A correspond in position, appearance and intensity to those obtained with solution B.

Rifampicin-related substances Carry out the test as described under 1.14.4 High-performance liquid chromatography, preparing the solutions and using the conditions given below under Assay method B.

Inject 20 µl of solution (4). The test is not valid unless the resolution between the peaks is at least 4.

Inject 20 µl each of solutions (1), (3) and (5).

In the chromatogram obtained with solution (5) the following impurity peaks are eluted at the following relative retention with reference to rifampicin (retention time about 25 minutes): rifampicin N-oxide about 0.3; 3-(isonicotinoylhydrazinomethyl) rifamycin [the "hydrazone" resulting from reaction between 3-formylrifamycin and isoniazid] about 0.5; rifampicin quinone about 0.7.

*Note: The means of identifying the impurity peaks is subject to confirmation.*
In the chromatogram obtained with solution (1), the area of any peak corresponding to the hydrazone impurity is not greater than the area of the principal peak in the chromatogram obtained with solution (3) (5.0%), the area of any peak corresponding to rifampicin quinone is not more than 0.8 times the area of the principal peak in the chromatogram obtained with solution (3) (4.0%) and the area of any other peak is not greater than 0.3 times the area of the principal peak in the chromatogram obtained with solution (3) (1.5%). The sum of the areas of all the peaks, other than the principal peak, is not greater than twice the area of the principal peak obtained with solution (3) (10.0% with reference to the content of rifampicin). Disregard any peak with an area less than 0.02 times the area of the principal peak in the chromatogram obtained with solution (3) (0.1%) and any peak with relative retention time less than 0.23 with reference to rifampicin.

Assay

A. For isoniazid, pyrazinamide and ethambutol hydrochloride Determine by 1.14.4 High-performance liquid chromatography, using a stainless steel column (15 cm x 4.6 mm) packed with particles of silica gel, the surface of which has been modified with chemically bonded octadecylsilyl groups, (5 µm)1. As the mobile phase, use a solution prepared as follows: dissolve 50 g ammonium acetate R and 0.2 g copper(II) acetate R in 1000 ml of water and adjust to pH 5.0 with glacial acetic acid R. Mix 940 ml of this solution with 60 ml methanol R. Prepare the following solutions in water. For solution (1) weigh and powder 20 tablets. Transfer a quantity of the powder equivalent to about 100 mg Ethambutol hydrochloride, accurately weighed, to a 500 ml volumetric flask. Dissolve in about 400 ml water by shaking for about 15 minutes. [If foaming occurs, use 400 ml of a 4% solution of methanol R in place of the water.] Dilute to 500 ml with water. Filter a portion of this solution through a 0.45 µm filter, discarding the first few ml of the filtered solution. For solution (2) dissolve 27.3 mg isoniazid RS, 145.5 mg pyrazinamide RS and 100 mg ethambutol hydrochloride RS in 500 ml water.

Operate with a flow rate of 2.0 ml per minute. As a detector use an ultraviolet spectrophotometer set at a wavelength of about 270 nm.

Inject 20 µl of solution (2). The assay is not valid unless the resolution between the isoniazid and pyrazinamide peaks is at least 2. (The peaks elute in the order, isoniazid, pyrazinamide and ethambutol hydrochloride.)

Inject alternately 20 µl each of solutions (1) and (2).

Measure the areas of the peak responses obtained in the chromatograms from solutions (1) and (2), and calculate the content of isoniazid, C₆H₇N₃O, pyrazinamide, C₅H₅N₃O and ethambutol hydrochloride, C₁₀H₂₄N₂O₂, 2HCl.

B. For rifampicin

Prepare fresh solutions and perform the assay without delay. Low-actininc glassware is recommended.

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1 Luna® is suitable
Determine by 1.14.4 High-performance liquid chromatography, using a stainless steel column (25 cm x 4.6 mm) packed with particles of silica gel, the surface of which has been modified with chemically bonded octadecylsilyl groups, (5 µm)². As the mobile phase, use a mixture of 6 volumes of methanol R and 4 volumes of phosphate buffer pH 7.0 (potassium dihydrogen phosphate R (0.01 mol/l), adjusted with sodium hydroxide (0.1 mol/l)) VS.

Prepare the following solutions in methanol R. For solution (1) weigh and powder 20 tablets. Without delay, shake a quantity of the powder equivalent to about 150 mg Rifampicin in 100 ml methanol R, filter and dilute 5 ml to 10 ml with methanol R. Solution (2) contains 0.75 mg rifampicin RS per ml. For solution (3) dilute a suitable volume of solution (1) to obtain a concentration equivalent to 37.5 µg Rifampicin per ml. Solution (4) contains 0.375 mg rifampicin RS per ml and 0.375 mg rifampicin quinone RS per ml. Solution (5) contains …mg of rifampicin impurity RS per ml. **Note: see under test for Rifampicin-related substance above**

Operate with a flow rate of 1.0 ml per minute. As a detector use an ultraviolet spectrophotometer set at a wavelength of about 254 nm.

Inject 20 µl of solution (4). The assay is not valid unless the resolution between the peaks is at least 4.

Inject alternately 20 µl each of solutions (1) and (2). Measure the areas of the peak responses obtained in the chromatograms from solutions (1) and (2), and calculate the content of rifampicin, C₄₃H₅₈N₄O₁₂.

**Dissolution test. To be added for rifampicin.**

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² Luna® is suitable